Scanning Probe Microscope Techniques for the Engineering of Nanoelectronic Devices

Landon Prisbrey,¹ Ji-Yong Park,² Kerstin Blank,³ Amir Moshar,⁴ and Ethan D. Minot¹

1. Department of Physics, Oregon State University, Corvallis, OR, USA. 2. Department of Physics and Division of Energy Systems Research, Ajou University, Suwon, Korea. 3. Institute for Molecules and Materials, Radboud University, Nijmegen, The Netherlands. 4. Asylum Research, Santa Barbara, CA, USA

BIOGRAPHY

Ethan D. Minot received his PhD in physics from Cornell University in 2004. He completed postdoctoral work at Delft University of Technology before joining the



physics faculty at Oregon State University. Dr Minot's research focuses on the electronic and optical properties of carbon nanotubes. Many of his experiments rely on scanning probe techniques such as atomic force microscopy and scanning photocurrent microscopy.

ABSTRACT

Using CNT network sensors as our working example, we review AFM-based techniques which are used to study and engineer nanoelectronic devices. We have used dc-EFM and ac-EFM to identify the locations and resistances of individual CNTs that are electrically connected in parallel. Next, SGM and tm-SGM were used to reveal the semiconducting response of each CNT. With the information available in these scans a single CNT with desirable properties was singled out for further experimentation. The unwanted CNTs were electrically cut with a biased AFM probe to leave a device containing a single CNT. An atomic-sized transistor with chemical functionality was engineered in the remaining CNT using a voltage pulse from the AFM probe. This nanotransistor was then demonstrated to be a single molecule sensor sensitive to EDC in an aqueous environment. This multi-step measurement and manipulation process illustrates the power of AFM-based techniques to map out and control the properties of nanoelectronic devices.

KEYWORDS

scanning probe microscopy, electric force microscopy, scanning gate microscopy, carbon nanotubes, nanoengineering, nanoelectronics

AUTHOR DETAILS

Dr Ethan D. Minot, Department of Physics, Oregon State University, Corvallis, Oregon 97331-6507, USA Tel: +1 541 737 9671 Email: minote@science.oregonstate.edu

Microscopy and Analysis 26(3):7-10 (AM), 2012

INTRODUCTION

Research in nanoscale systems relies heavily on scanning probe microscopy. Many of these nanoscale systems have unique properties that are amenable to scanning probe techniques and, therefore, a wide variety of specialized scanning probe techniques have been developed. For example, in the field of magnetic memory research, magnetized probes are used to map magnetization patterns [1], while ferroelectric materials are studied by locating the electric fields from polarized domains [2]. In many cases the scanning probe microscope is used both as a manipulation tool as well as an imaging tool. It is possible, for example, to flip discrete magnetic or ferroelectric domains and then image the resulting changes [2, 3].

Nanoscale Electronic Devices

A growing toolkit of scanning probe microscopy-based techniques is enabling new ways to build and investigate nanoscale electronic devices. In this article we review key techniques that allow for mapping the distribution of electrical resistances, measuring local field-effect sensitivity, and engineering new electrical characteristics into nanoscale devices.

Here we review several advanced techniques to characterize and manipulate nanoelectronic devices using an atomic force microscope (AFM). Starting from a carbon nanotube (CNT) network device that is fabricated by conventional photolithography (micrometer-scale resolution), individual carbon nanotubes can be characterized, unwanted carbon nanotubes can be cut, and an atomic-sized transistor with single molecule detection capabilities can be created. While we focus on CNT devices, many of the same techniques are being used for nanowire [4] and graphene [5] devices.

These measurement and manipulation processes were performed with the MFP-3D AFM (Asylum Research, Santa Barbara, CA) with the Probe Station Option (Figure 1) and illustrate the recent progress in AFM-based techniques for nanoelectronics research.

We first discuss how electric force microscopy (EFM) and scanning gate microscopy (SGM) techniques are used to characterize the electrical response of nanoelectronic devices. The information contained in EFM and SGM images is extremely valuable for relating the local electrical behavior of a device to the global device properties. In the specific example of our CNT network device, we use the knowledge of local resistance and local semiconducting behavior to choose the 'best' CNT out of a mixture of parallel CNTs.

ELECTRIC FORCE AND SCANNING GATE MICROSCOPY

To acquire EFM and SGM signals, a conducting AFM probe hovers above an electrically contacted device (Figure 2a). During a 'hover pass', the microscope maintains a constant separation between the conductive AFM tip and the surface (typical separations are 20 to 200 nm). To achieve this constant separation distance, the microscope alternates between standard topographical line scans and hover pass line scans. A typical electrical contact configuration involves a current amplifier and three voltage sources (V_{sd} , V_{tip} , and V_{bg}). Depending on the particular imaging technique, different voltage signals will be applied and the AFM tip may or may not be piezoelectrically driven (see Table 1).

To demonstrate the utility of EFM and SGM,



Figure 1:

Experimental setup. The Probe Station attaches to the MFP-3D AFM scanner and allows easy electrical probing of sample properties, electrical biasing, and other measurements while the sample is being scanned with the AFM. A variety of electrical connections can be made and combined with various imaging modes. we used dc-EFM [6], ac-EFM [7], SGM [8] and tip modulated SGM (tm-SGM) [9] to image a carbon nanotube (CNT) network device (Figure 2). The device was fabricated by growing CNTs on a SiO₂/Si substrate (300 nm oxide) [10], and then patterning electrodes with photolithography. The images in Figures 2b and 3a are a combination of a dc-EFM data (color scale) and AFM topography. Many individual CNTs are seen bridging the gap between the source and drain electrodes. We are interested in learning which CNTs are metallic, which are semiconducting, which are well connected to the metal electrodes and which contain natural defects.

Electric Force Microscopy Techniques

The electrostatic forces which give imaging contrast in dc-EFM [6] (Figures 2b and 3a) are due to a voltage difference between the tip and the grounded electrodes. During each hover pass, the AFM cantilever was piezoelectrically driven at a fixed frequency (the fundamental resonance of the cantilever). If the AFM tip is above a grounded conductor, a vertical gradient in electrostatic force causes a shift in the resonance frequency of the cantilever. This resonant frequency shift modulates both the phase and amplitude of the cantilever motion. The dc-EFM image shown in Figure 3a allows us to quickly locate all conducting objects in the large (60 µm) imaging area and requires minimal setup.

The vertical gradient in electrostatic force, which gives rise to the dc-EFM signal, depends on the tip-sample capacitance and the tip-sample voltage difference. When the tip hovers above a large-area electrode, the vertical gradient in tip-sample capacitance is large. When the tip hovers over a small CNT, the vertical gradient in tip-sample capacitance is small. Thus, the EFM signal from CNTs is weaker than from the electrodes. There has been steady progress in mapping out the vertical gradient in tip-sample capacitance to allow more quantitative imaging of the potential difference between the tip and the sample [11,12].

Figure 3b shows an ac-EFM [7] image in which different voltage signals are sent to the source and drain electrodes, respectively. This type of ac-EFM scan is used to visualize the voltage profile along the length of electricallybiased CNTs. A similar measurement can be done with dc-EFM, however, the ac-EFM signal is cleaner (more discussion below). CNTs labeled 1-4 in Figure 3b exhibit a sharp voltage drop where they meet the inner electrode (the length of the CNT appears uniform black). This sudden voltage drop indicates a high contact resistance. The CNT labeled 5 exhibits a sharp voltage drop midway along its length. The location of the voltage drop corresponds to a sharp bend in the CNT, showing that the bend is a point of high resistance. CNTs labeled 6-10 show a gradual voltage gradient along their length, indicating that these CNTs are well connected to both electrodes, and are free from major electrical defects. These CNTs with small contact resistance will be subsequently singled out for additional engineering and experiments.



Figure 2:

(a) Schematic illustrating a carbon nanotube field-effect transistor and 'hover pass' (see text) microscopy geometries. Not to scale. (b) AFM topography colored with EFM phase ($V_{tin} = 8 V$; height = 20 nm).

	$V_{\sf sd}$	Piezoelectrically Driven?	'Hover Pass' Signal
dc-EFM	DC	yes	phase and amplitude
ac-EFM	$AC \mathbf{@} f_{cantilever}$	no	amplitude
SGM	DC	no	$I_{\rm sd}$
tm-SGM	DC	yes	$I_{\rm sd}\text{, AC @}f_{\rm cantilever}$

Table 1:

Key differences between modes used for imaging nanoelectronic devices.



Figure 3:

'Hover pass' microscopy data of a typical carbon nanotube field-effect transistor. (a) EFM ($V_{sd} = 20 \text{ mV}$, $V_{bg} = 0 \text{ V}$, $V_{tip} = 5 \text{ V}$, height = 20 nm). Color scale shows phase of cantilever response. (b) Alternating current EFM ($V_{sd} = 100 \text{ mV} @ 63 \text{ kHz}$, $V_{bg} = 0 \text{ V}$, $V_{tip} = 2 \text{ V}$, height = 20 nm). Color scale shows amplitude of cantilever response. (c) SGM ($V_{sd} = 20 \text{ mV}$, $V_{bg} = 0 \text{ V}$, $V_{tip} = 5 \text{ V}$, height = 20 nm). Color scale shows the change in current Δ I_{sd} through the device. (d) Tip-modulated SGM ($V_{sd} = 20 \text{ mV}$, $V_{bg} = 0 \text{ V}$, $V_{tip} = 5 \text{ V}$, height = 20 nm). Color scale shows the change in current at the cantilever fundamental frequency, Δ I_{sd}, ac.

NANOELECTRONICS

To obtain the ac-EFM signal in Figure 3b, an AC bias was applied to the outer electrode (frequency matching the cantilever's mechanical resonance) while the inner electrode was grounded and the AFM tip was biased to 2 V. During the hover pass the cantilever is not piezoelectrically driven, instead cantilever oscillations are driven by capacitive coupling to the AC biased electrode. These oscillations are detected by monitoring the cantilever deflection signal using a lock-in amplifier. The oscillation amplitude is plotted as a function of tip position to create the ac-EFM image. Cantilever oscillations are strongest when the probe is near the biased electrode. The ac-EFM signal is 'cleaner' than dc-EFM signals because stray DC signals (such as charge on the insulating substrate) are invisible in ac-EFM.

Scanning Gate Microscopy Techniques

We now turn our attention to scanning gate techniques, which allow researchers to further characterize the electronic properties of individual CNTs. Scanning gate techniques identify the local semiconducting response of a nanoscale conductor, revealing the presence of localized semiconducting 'hotspots' and bottlenecks for electron transport [8].

Figure 3c shows an SGM image, where the AFM probe acts as a roaming gate electrode to modulate the doping in the underlying electronics. The SGM image (Figure 3c) shows a single feature, indicating that the global semiconductor response of the device is dominated by a single CNT (labeled 10 in Figure 3b). Other CNTs are present in the imaging area, but the SGM signals from these are significantly smaller. Methods to improve sensitivity are discussed below.

The imaging contrast in Figure 3c corresponds to changes in global device current when the biased probe locally changes the doping level in the underlying electronics. A small source-drain voltage (V_{sd}) is applied across the device to drive global current. By monitoring changes in current as a function of tip position, the local field-effect sensitivity can be mapped.

A variation of SGM which offers greater sensitivity is tip-modulated SGM (tm-SGM) [9]. Compared to conventional SGM, tm-SGM offers higher spatial resolution and is capable of resolving weaker signals. Figure 3d shows a tm-SGM image. This scan reveals five CNTs with a semiconducting response (labeled 6-10 in Figure 3b). One CNT contains a region of highly localized semiconducting response, indicating the presence of a natural defect (labeled 8 in Figure 3b).

In tm-SGM the AFM cantilever is biased and piezoelectrically driven at its resonant frequency, resulting in an AC electric field which modulates the local doping level in the underlying electronics. The local field-effect response of the sample is mapped as a function of tip position by isolating the component of the conductance signal at the cantilever resonant frequency using a lock-in amplifier.

Interpreting SGM and EFM Data

The detailed characterization outlined in



Figure 4:

Electrical cutting of unwanted carbon nanotubes. (a) Electric force microscope image of a device. Cutting path indicated by dotted arrow ($V_{sd} = 25 \text{ mV}$, $V_{ba} = 0 \text{ V}$, $V_{tin} = 8 \text{ V}$, height = 20 nm). (b) Time trace of cutting event ($V_{sd} = 25 \text{ mV}$, $V_{ba} = 0 \text{ V}$, $V_{tin} = -8 \text{ V}$, contact force = 6-9 nN).



Figure 5:

Defect engineering in a carbon nanotube field-effect transistor. (a) Schematic illustrating defect engineering geometry. (b) Transistor characteristics before (dashed black) and after (solid red) defect creation. The red curve has been multiplied by 10 ($V_{sd} = 25 \text{ mV}$). (c) AFM topography of a CNT device. A defect was engineered at the location indicated by an arrow. (d) SGM data of the device shown in (c) taken before and (e) after defect engineering ($V_{sd} = 25 \text{ mV}$). $V_{tip} = 0 \text{ V}$, $V_{tip} = 8 \text{ V}$, height = 20 nm). The color scale ($\Delta I_{sd} I_0$) indicates changes in device current relative to the baseline value, I_0 in (d) and (e) the values of I_0 are 280 nA and 7 nA, respectively.

Figure 3 allows us to pick the best CNT out of the ten CNTs for building a nanotransistor with chemical functionality. Using dc-EFM (Figure 3a), we identified the locations of all CNTs in a large area scan, providing a road map for engineering. With ac-EFM (Figure 3b) we identified desirable CNTs with low contact resistance (no sudden drop in voltage at metal-CNT contact) and an absence of natural defects (no sudden drop in voltage along length of CNT). Finally, using SGM methods (Figure 3 c,d), we identified CNTs with a semiconducting response.

The most desirable CNT for building a nanotransistor is one with low contact resistance, no natural defects, and a uniform semiconducting response along its length. The CNT labeled 10 in Figure 3b satisfies these criteria, and is singled out for further experimentation.

Atomic Force Microscopy Engineering

With initial characterization complete, we move to AFM manipulation techniques. Once a desired CNT is identified, we use AFM engineering to prune unwanted CNTs [13], then engineer an atomic-sized transistor element into one remaining CNT [14]. These techniques combine AFM lithography modes with control of the tip-sample voltage. Mechanical manipulation by AFM has also been used by many authors [15, 16], but is not discussed here.

To cut unwanted CNTs, a Pt-coated AFM probe is biased to $V_{\rm tip}$ = -10 V, engaged with the surface, and then dragged through the CNTs in contact mode (contact force ~6-9 nN). Previously obtained dc-EFM and ac-EFM scans, such as in Figure 3 a,b, provide a convenient road map of the locations of the unwanted CNTs. Figure 4 shows a time trace of global device conductance during electrical cutting. Sharp decreases in device conductance are observed as the CNTs are severed. Afterwards, ac-EFM and SGM scans may be used to verify that a single CNT forms the only electrical contact between the electrodes.

A single CNT can be converted into an atomic-sized transistor by further AFM-based engineering. Such nanotransistors approach the ultimate limit of miniaturization. To engineer an atomic-sized transistor, a conducting AFM probe is brought into gentle contact with the CNT and a -3 V, 15 ms square wave pulse is applied to the tip (Figure 5a). During the pulse, a chemical defect is incorporated into the sidewall of the CNT, and the device's electrical properties change significantly as a result (Figure 5b) [14, 17, 20]. Typical defects add between 10 k Ω to 1 M Ω to the overall resistance of the circuit, and change the field-effect sensitivity of the device.

Figure 5c shows a single-CNT device produced by the electrical 'nicking' method described above. SGM measurements show that the pristine CNT is uniformly gate sensitive (Figure 5d). Figure 5e shows an identical measurement following defect engineering where gate sensitivity is localized to the region around the defect. The SGM scan reveals that the defect acts as a gate sensitive bottleneck for transport, and the remainder of the CNT serves only as contact electrodes to the miniature transistor.



ΠΠ

Single-molecule detection of EDC.

(a) Schematic illustrating detection geometry. The liquid potential is controlled with a liquid-gate electrode biased to V_{ler} .

(b) I_{sd} trace of a nanotransistor in MES buffer (pH 4.5, V_{sd} = 25 mV, V_{lg} = 250 mV). (c) I_{sd} trace of the same device following the addition of 20 μ M EDC (pH 4.5, V_{sd} = 25 mV, V_{lg} = 100 mV).

A unique attribute of point-defect nanotransistors is their sensitivity to electrostatic potential within a very small detection volume. As such, they are ideally suited for singlemolecule sensing applications. Recently, pointdefect nanotransistor sensors have been used to study biochemical reactions at the single molecule level [18, 19]

Figure 6 demonstrates the use of an AFMengineered point-defect nanotransistor as a single-molecule sensor. The reaction between N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide (EDC) and a carboxyl group is used as a model reaction. EDC is frequently used to activate carboxyl groups in bioconjugation chemistry, and reacts reversibly with a carboxyl point defect on the CNT sidewall. The pointdefect nanotransistor was immersed in an electrolyte solution containing EDC (Figure 6a). Figures 6 b,c show the conductance data for a point-defect CNT before and after the addition of 20 µM EDC, respectively. Discrete switching events are observed in the presence of EDC.

This two-state telegraph noise is believed to reflect the fluctuating electrostatic environment as individual EDC molecules react with the carboxyl group on the CNT sidewall [18].

CONCLUSIONS

Using CNT network sensors as our working example, we have reviewed AFM-based techniques which are used to study and engineer nanoelectronic devices. We have used dc-EFM and ac-EFM to identify the locations and resistances of individual CNTs that are electrically connected in parallel. Next, SGM and tm-SGM were used to reveal the semiconducting response of each CNT. With the information available in these scans a single CNT with desirable properties was singled out for further experimentation. The unwanted CNTs were electrically cut with a biased AFM probe to leave a device containing a single CNT. An atomic-sized transistor with chemical functionality was engineered in the remaining CNT using a voltage pulse from the AFM probe. This nanotransistor was then demonstrated to be a single molecule sensor sensitive to EDC in an aqueous environment. This multi-step measurement and manipulation process illustrates the power of AFM-based techniques to map out and control the properties of nanoelectronic devices.

ACKNOWLEDGEMENTS

We thank Kristina Prisbrey for assistance with Figures 2, 5, and 6, and Bo Criss for assistance with Figure 3. We thank Sophie Ripp for initial experiments with EDC using suspended CNTs. We thank Paul Schuele and Sharp Labs of America for assistance with device fabrication. This work is funded by the Human Frontier Science Program.

REFERENCES

- 1. Parkin, S. et al. Science 320:190-194, 2008.
- Gruverman, A. J. Vacuum Sci. & Technology B: Microelectronics and Nanometer Structures 14:602, 1996.
- Gibson, G. A., Smyth, J. F., Schultz, S., Kern, D. P. IEEE Transactions on Magnetics 27:5187-5189, 1991.
- Zhou, X., Dayeh, S. A., Wang, D., Yu, E. T. Applied Physics Letters 90:233118, 2007.
- 5. Jalilian, R. et al. Nanotechnology 22:295705, 2011.
- Martin, Y., Abraham, D. W., Wickramasinghe, H. K. Applied Physics Letters 52:1103, 1988.
- Bachtold, A., Fuhrer, M. S., Plyasunov, S. et al. Phys. Rev. Lett. 84:6082, 2000.
- 8. Tans, S. J., Dekker, C. Nature 404:834-835, 2000.
- 9. Wilson, N. R., Cobden, D. H. Nano Letters 8:2161-2165, 2008.
- 10. Kong, J. et al. Nature 395:878-881, 1998.
- 11. Lei, C. H. et al. Nanotechnology 15:627-634, 2004.
- Yan, M., Bernstein, G. H. Surface and Interface Analysis 39:354-358, 2007.
- 13. Park, J.-Y. et al. Appl. Phys. Lett. 80:4446-4448, 2002.
 - 14. Park, J.-Y. Appl. Phys. Lett. 90:023112-3, 2007.
 - Junno, T., Deppert, K., Montelius, L., Samuelson, L. Applied Physics Letters 66:3627, 1995.
 - Postma, H. W. C., Teepen, T., Yao, Z., Grifoni, M., Dekker, C. Science 293:76-79, 2001.
 - 17. Kim, D.-H., Koo, J.-Y., Kim, J.-J. Phys. Rev. B 68:113406, 2003.
 - 18. Goldsmith, B. R., Coroneus, J. G., Kane, A. A., Weiss, G. A.,
- Collins, P. G. Nano Letters 8:189-194, 2008.
- 19. Sorgenfrei, S. et al. Nature Nano. 6:126-132, 2011.
- 20. Prisbrey, L. et al. J. Phys. Chem. C 116:1961-1965, 2012.

©2012 John Wiley & Sons, Ltd